- [2] Knudsen TB, Gray MK, Church JK, et al. Early postimplantation embryolethality in mice following in utero inhibition of adenosine deaminase with 2'-deoxycoformycin. Teratology 1989;40:615-26.
- [3] Blackburn MR, Datta SK, Kellems RE. Adenosine deaminase—deficient mice generated using a two stage genetic engineering strategy exhibit a combined immunodeficiency. J Biol Chem 1998;273:5093-100.
- [4] Yoneyama Y, Sawa R, Suzuki S, et al. Regulation of plasma adenosine levels in normal pregnancy. Gynecol Obstet Invest 2002;53:71-4.
- [5] Battistuzzi G, Iudicone P, Santolamazza P, Petrucci R. Activity of adenosine deaminase allelic forms in intact erythrocytes and in lymphocytes. Ann Hum Genet 1981;45:15-9.
- [6] Bottini E, Carapella E, Cataldi L, et al. Adenosine deaminase polymorphism. Associations at clinical level suggest a role in cell functions and immune reactions. J Med Genet 1981;18:331-4.
- [7] Nicotra M, Bottini N, Grasso M, et al. Adenosine deaminase and human reproduction: a comparative study of fertile women and women with recurrent spontaneous abortion. Am J Reprod Immunol 1998;39:266-70.
- [8] Borgiani P, Gloria-Bottini F, Lucarini N, et al. Diabetic pregnancy: is there intrauterine selection of ADA polymorphism? Am J Hum Genet 1991:49:464-6.
- [9] Gloria-Bottini F, Pietropolli A, Coppeta L, Magrini A, Bergamaschi A, Bottini E. The genetics of feto-placental development: a study of acid phosphatase locus 1 and adenosine deaminase polymorphisms in a consecutive series of newborn infants. Reprod Biol Endocrinol 2008;6:38.

Reply: Purine metabolite levels in preovulatory human follicles may hold the key to ovarian hyperstimulation syndrome

To the Editor:

We are in agreement with Valerio Napolioni that adenosine's importance in reproductive biology may be wider and more variable than the association with energy charge that we describe. Indeed, we feel that the potent pharmacology of this purine may be unwelcome under emerging circumstances. Tremendous progress has been made in assisted reproductive technology since the birth of the first baby in 1978 by in vitro fertilization (IVF). The development of intracytoplasmic sperm injection in the early 1990s further increased the pregnancy and live birth rates [1]; nonetheless, a serious iatrogenic illness arose from this technology in the form of ovarian hyperstimulation syndrome (OHSS). This is experienced by approximately 5% to 10% of women undergoing IVF; and the clinical symptoms of OHSS are graded mild, moderate, and severe. Mild symptoms include abdominal bloating and feeling of fullness, nausea, diarrhea, and slight weight gain. The progression to moderate symptoms is defined by excessive weight gain (weight gain of >2 lb/d), increased abdominal girth, vomiting, diarrhea, concentrated urine, and excessive thirst. Severe symptoms are marked abdominal distension due to ascites, pulmonary edema, and chest pain [2].

The molecular cause of OHSS has been put down to a soluble factor found to be produced by multiple follicles that arise as a result of deliberate ovarian stimulation. Research into follicular fluid has been undertaken in many different ways: immunoassays for specific molecules or hormones [3-5], proteomic studies by 2-dimensional electrophoresis

and mass spectrometry analysis [6], and granulosa cell messenger RNA quantification for inhibin-activin-follistatin system by polymerase chain reaction [7]. The primary focus in the search for the molecular agent responsible for OHSS has centered on vascular endothelial growth factor (VEGF), perhaps because of our current preoccupation with genes and proteins rather than smaller potent bioactive molecules. There is a presumption that VEGF levels are supraphysiologic in follicular fluid and will cause local blood vessels to become leaky [2]. Unfortunately, VEGF levels are not particularly elevated in follicular fluid compared with other sources [8]. However, smaller vasodilatory purine metabolites are present in follicular fluid in abundance [9]. In the 1980s, Downs et al [10] studied their roles as meiotic inhibitors (in mice predominantly). Later, Lavy et al [11], studying purine metabolite levels in human follicles from both natural and stimulated cycles, claimed that adenosine was the inhibitor of human oocyte maturation. In our recent study, we found that hypoxanthine levels were extremely variable, but adenosine was a consistent component, and levels were supraphysiologic—the smaller follicles contained a much greater concentration of adenosine than the larger ones [9].

Adenosine's other biological actions make it a significant contender as the molecular cause of OHSS: adenosine is a powerful vasodilator; and when administered by intravenous infusion, it can produce substantial hypotension. Acting via adenosine A2 receptors, it induces smooth muscle relaxation, especially in the coronary circulation; but because of its extremely rapid metabolism, it is very short acting. It is common practice to infuse adenosine into coronary arteries when imaging occlusions, but it is not used clinically as a vasodilator. However, in such patients, most of adenosine's side effects are related to its vasodilatory properties. Furthermore, peripheral microvascular endothelia local production of vascular permeability factor/VEGF A is upregulated 2- to 3-fold by adenosine [12].

A wave of symptoms due to vasodilation and increased vessel permeability spreading from the ovaries to the abdomen and on to the lungs would be consistent with unopposed peritoneal infusion of adenosine (leaking from multiple follicles). This is prevented by adenosine deaminase, or ADA (also referred to as adenosine aminohydrolase, EC 3.5.4.4), a ubiquitous enzyme that appears to be particularly important in the development of thymocytes. ADA converts adenosine into inosine through the hydrolysis of the purine amino group, with an estimated half-life of 1 second (Fig. 1). ADA is present in all tissues, but activity is particularly high in thymocytes of the thymic cortex. There are 2 enzymes that carry out ADA activity, called ADA1 and ADA2. ADA1 a 40-kd monomeric protein with a 200-kd noncatalytic combining protein, and it is responsible for about 90% of adenosine deamination. ADA2 is somewhat larger at 110 kd and appears to play a general adenosine deamination role in serum. Total absence of ADA activity results in a form of severe combined immunodeficiency.

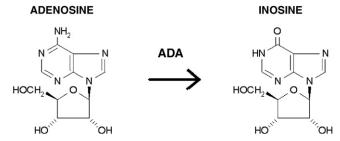


Fig. 1. Adenosine is deactivated by its rapid conversion to inosine through the hydrolysis of the purine amino group by ADA.

However, as pointed out by Valerio Napolioni, polymorphic variants have now been found with reduced rates of catalytic activity. In some circumstances, this is positively advantageous. Adenosine is released by cardiomyocytes in response to ischemia and is cardioprotective in this regard [13]. Genotypic variants resulting in reduced metabolism of, or increased receptor response to, adenosine result in a phenotypic group more likely to survive ischemic events [14,15]. The question to be investigated is this: do these same genotypes result in an IVF patient phenotype prone to the development of OHSS?

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References

- Palermo G, Joris H, Devroey P, Van Steirteghem AC. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. Lancet 1992;340:17-8.
- [2] Delvigne A, Rozenberg S. Review of clinical course and treatment of ovarian hyperstimulation syndrome (OHSS). Hum Reprod Update 2003;9:77-96.
- [3] Sabatini L, Wilson C, Lower A, Al-Shawaf T, Grudzinskas G. Superoxide dismutase activity in human follicular fluid after controlled ovarian hyperstimulation in women undergoing in vitro fertilisation. Fertil Steril 1999;72:1027-34.
- [4] Manau D, Balasch J, Jimenez W, Fabregues F, Civico S, Casamitjana R, et al. Follicular fluid concentrations of adrenomedulllin, vascular endothelial growth factor and nitric oxide in IVF cycles: relationship to ovarian response. Hum Reprod 2000;15:1295-9.
- [5] Mendoza C, Ruiz-Requena E, Ortega E, Cremades N, Martinez F, Bernabeu R, et al. Follicular fluid markers of oocyte developmental potential. Hum Reprod 2002;17:1017-22.
- [6] Anahory T, Dechaud H, Bennes R, Marin P, Lamb NJ, Laoudj D. Identification of new proteins in follicular fluid of mature human follicles. Electrophoresis 2002;23:1197-202.
- [7] Fujiwara T, Lambert-Messerlian G, Sidis Y, Leykin L, Isaacson K, Toth T, et al. Analysis of follicular fluid hormone concentrations and granulosa cell mRNA levels for the inhibin-activin-follistatin system:

- relation to oocyte and embryo characteristics. Fertil Steril 2000;74:348-55.
- [8] Tozer AJ, Iles RK, Iammarrone E, Gillott CM, Al-Shawaf T, Grudzinskas JG. The effects of 'coasting' on follicular fluid concentrations of vascular endothelial growth factor in women at risk of developing ovarian hyperstimulation syndrome. Hum Reprod 2004:19:522-8.
- [9] Wen X, Perrett D, Jones N, Tozer AJ, Docherty SM, Iles RK. High follicular fluid adenosine levels may be pivotal in the metabolism and recycling of adenosine nucleotides in the human follicle. Metabolism 2009 [Epub ahead of print] PubMed PMID: 20045541.
- [10] Downs S, Daniel S, Bornslaeger E, Hoppe P, Eppig JJ. Maintenance of meiotic arrest in mouse oocytes by purines: modulation of cAMP levels and cAMP phosphodiesterase activity. Gamete Res 1989;23:323-34.
- [11] Lavy G, Beheman HR, Polan ML. Purine levels and metabolism in human follicular fluid. Hum Reprod 1990;5:529-32.
- [12] Fischer S, Sharma HS, Karliczek GF, Schaper W. Expression of vascular permeability factor/vascular endothelial growth factor in pig cerebral microvascular endothelial cells and its upregulation by adenosine. Brain Res Mol Brain Res 1995;28:141-8.
- [13] Asakura M, Asanuma H, Kim J, Liao Y, Nakamaru K, Fujita M, et al. Impact of adenosine receptor signaling and metabolism on pathophysiology in patients with chronic heart failure. Hypertens Res 2007;30:781-7.
- [14] Safranow K, Czyzycka E, Binczak-Kuleta A, Rzeuski R, Skowronek J, Wojtarowicz A, et al. Association of C34T AMPD1 gene polymorphism with features of metabolic syndrome in patients with coronary artery disease or heart failure. Scand J Clin Lab Invest 2009;69:102-12.
- [15] Hand BD, Roth SM, Roltsch MH, Park JJ, Kostek MC, Ferrell RE, et al. AMPD1 gene polymorphism and the vasodilatory response to ischemia. Life Sci 2006;79:1413-8.

How the "A" to "C" conversion may create a new splice acceptor site?

To the Editor:

We have read with great interest the article by Katsumata et al entitled "Novel intronic *CYP21A2* mutation in a Japanese patient with classic salt-wasting steroid 21-hydroxylase deficiency" [1].

The authors report a novel *CYP21A2* mutation (IVS-9A>C) at -9 position of intron 9 in a Japanese male patient suffering from a severe classic form of congenital adrenal hyperplasia.

Katsumata et al perform complex in vitro experiments showing that the transient expression of the *CYP21A2* IVS-9A>C mutation in COS-1 cells creates an aberrant splice acceptor site at -7 position of intron 9, inactivating the original slice acceptor site. The result is the complete deficiency of 21-hydroxylase activity and loss of immunoreactive CYP21A2 protein.

In line with the reported data, we now ask ourselves how the "A" to "C" conversion at -9 position of *CYP21A2* intron 9 may create a new splice acceptor site. Generally, a canonical acceptor splice site is "AG"; and only this consensus bases sequencing can be correctly recognized by splicing factors [2].